#### MASTER OF SCIENCE (BIOTECHNOLOGY) TWO-YEAR FULL-TIME PROGRAMME

#### AFFILIATION

The proposed programme shall be governed by the Department of Biotechnology, Faculty of Science, D.D.U. Gorakhpur University, Gorakhpur–273009.

#### **PROGRAMME STRUCTURE**

The M.Sc.Programme is divided into two Parts as under. Each part will consist of two Semesters as given below.

		Semester – Odd	Semester – Even
Part I	First Year	Semester – 1	Semester $-2$
Part II	Second Year	Semester – 3	Semester – 4

For semester-1 and 2 would consist of four theory papers of 4 credits each, one laboratory course of 8 credits and one seminar of 1 credit. In semester 3 one elective course of 4 credits would also be offered for other Departments of Faculty of Sciences besides four theory papers (core courses), one laboratory course and seminar. In semester 4, students have to select three out of four papers (elective) along with dissertation (core) and one seminar of 1 credit. **Dissertation/project work will be carried out by students under supervision of faculty members.** 

The schedule of papers prescribed for various semesters shall be as follows:

#### PART I: Semester - 1

IANI	1: Semester – 1	
1	BT 501	Microbiology
2	BT 502	Molecular Biology
3	BT 503	Biochemistry
4	BT 504	Cell Biology
5	BT 505	Laboratory course-I
6	BT 506	Seminar
PART	I: Semester – 2	
1	BT 507	Recombinant DNA technology
2	BT 508	Enzyme Technology
3	BT 509	Immunology
4	BT 510	Bioinformatics, Biostatistics and Genomics
5	BT 511	Laboratory course-II
6	BT 512	Seminar

#### PART II: Semester – 3

	III. Demester	
1	BT 513	Plant Biotechnology
2	BT 514	Animal Biotechnology
3	BT 515	Bioprocess Technology
4	BT 516	Environmental Biotechnology
5	BT 517	Introductory Biotechnology (Elective for other department)
6	BT 518	Laboratory course-III
7	BT 519	Seminar

**PART II: Semester – 4 (Elective course)** 

- Students will select three papers (courses) out of four papers from MBT-401, MBT-402, MBT-403 and MBT-404.
- Each student will have to submit an allotted dissertation (MBT-406), which would be based on research works and will submit a report on which Viva-voce will be conducted.

1	BT 520	Genomics for crop improvement(Elective)
2	BT 521	Proteomics and Nanobiotechnology (Elective)
3	BT 522	Biosafety, IPR and Bioethics (Elective)
4	BT 523	Animal Cell and Tissue Culture (Elective)
5	BT 524	Seminar
6	BT 525	Dissertation

M. Sc. Biotechnology	М.	Sc. Bio	otechno	logy
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Course No.	Course Title	Туре	Credit
		(Core/Elective)	
<b>DE 5</b> 04	Semester 1	~	
BT-501	MICROBIOLOGY	Core	4+0
BT-502	MOLECULAR BIOLOGY	Core	4+0
BT-503	BIOCHEMISTRY	Core	4+0
BT-504	CELL BIOLOGY	Core	4+0
BT-505	LABORATORY COURSE-I (Based on MBT-101, 102, 103 & 104)		0+8
BT-506	SEMINAR		0+1
<b>DI</b> 500		Total	25
	Semester 2	Total	20
BT 507	RECOMBINANT DNA TECHNOLOGY	Core	4+0
BT 508	ENZYME TECHNOLOGY	Core	4+0
BT 509	IMMUNOLOGY	Core	4+0 4+0
BT 510	BIOINFORMATICS, BIOSTATISTICS	Core	4+0 4+0
DI 510	AND GENOMICS		┓┰╻
BT 511	LABORATORY COURSE-II (Based on		0+8
<b>DI</b> 311	MBT-201, 202, 203 & 204)		0-0
BT 512	SEMINAR		0+1
DI 312	SEMINAR	Total	25
	Semester 3	Total	23
BT 513	PLANT BIOTECHNOLOGY	Core	4+0
BT 514	ANIMAL BIOTECHNOLOGY	Core	4+0 4+0
BT 515	BIOPROCESS TECHNOLOGY	Core	4+0 4+0
BT 516	ENVIRONMENTAL BIOTECHNOLOGY	Core	4+0 4+0
BT 517	INTRODUCTORY BIOTECHNOLOGY	Elective for	4+0 4+0
DI 317	INTRODUCTORT BIOTECHNOLOOT	other	4+0
BT 518	LABORATORY COURSE-III (Based on	department)	0+8
<b>DI 010</b>	MBT-301, 302, 303 & 304)		
BT 519	Seminar		0+1
		Total	25
	Semester 4		
BT 520	GENOMICS FOR CROP IMPROVEMENT	Elective	4+0
BT 521	PROTEOMICS AND	Elective	4+0
	NANOBIOTECHNOLOGY		
BT 522	BIOSAFETY, IPR AND BIOETHICS	Elective	4+0
BT 523	ANIMAL CELL AND TISSUE CULTURE	Elective	4+0
BT 524	SEMINAR	Core	0+1
BT 525	DISSERTATION	Core	0+12
		Total	25
<b>GRAND TOT</b>			100

# Semester-1

BT-501: MICROBIOLOGY	4+0
(Core)	
THEORY	

### Unit-1:

Microbial diversity and systematics, Modern approaches to bacterial taxonomy, polyphasic classification, General characteristics of primary domains and of taxonomic groups belonging to Bacteria, Archaea and Eukarya, Nomenclature and outline of bacterial classification as per Bergey's Manual, Accessing microbial diversity using molecular methods such as Denaturing Gradient Gel Electrophoresis (DGGE), Temperature Gradient Gel Electrophoresis (TGGE), Amplified rRNA restriction analysis, terminal Restriction Fragment Length Polymorphism (T-RFLP), 16S rDNA sequencing, metagenomics.

#### Unit-2:

Methods in Microbiology: Theory and practice of sterilization, Pure culture techniques, Principles of microbial nutrition, Construction of culture media, Enrichment culture techniques, Isolation and culture of aerobic and anaerobic bacteria, Culture collection, preservation and maintenance of microbial cultures.

#### Unit-3:

Metabolic Diversity among Microorganism: Microbial Nutrition: nutritional types and modes of nutrition in bacteria, Extremophiles. Microbial growth: The definition of growth, mathematical expression of growth, growth curve, measurement of growth and growth yields; Synchronous growth and Continuous culture.

# Unit-4:

Chemotherapy/Antibiotics Antimicrobial agents; Antibiotics: Penicillins and Cephalosporins and Broad- spectrum antibiotics, sulfa drugs, Antifungal antibiotics, Mode of action, Molecular mechanism of drug resistance. Bacterial Genetic System: Transformation, Conjugation, Transduction, Recombination, bacterial genetic map with reference to *E coli*.

- Brock Biology of Microorganisms, 9<sup>th</sup> Edition. By *Michael T. Madigan, John M. Martinko, Jack Parker*. Prentice Hall, Inc.
- Microbiology, 4<sup>th</sup> Edition. By *Lansing M. Prescott, John P. Harley, Donald A. Klein*. WCB McGraw Hill.
- General Microbiology, 5<sup>th</sup> Edition by *Roger Y. Stanier, John L. Ingraham, Mark L. Wheelis, Page R. Painter*, Macmillan Press Limited.
- Microbiology: Principles and Explorations, 5<sup>th</sup> Edition. By *Jacquelyn G. Black*, John Wiley& Son, Inc.

Prokaryotic and eukaryotic genome organization, structural elements of chromosome and construction of artificial chromosome. DNA replication: Enzymes, accessory proteins and mechanisms of prokaryotic and eukaryotic DNA replication.

# Unit 2:

Fine structure of gene, molecular basis of spontaneous and induced mutations and their role in evolution; DNA damage and repair, DNA amplification and rearrangement. Antisense and Ribozyme Technology: Inhibition of splicing, polyadenylation and translation, disruption of RNA structure and capping, Biochemistry of ribozyme, hammerhead, hairpin and other ribozymes, applications of anti-sense and ribozyme technologies.

# Unit 3:

Transcription: Organization of transcriptional units, mechanisms of transcription and its regulation in prokaryotes and eukaryotes, Operon concept, attenuation and antitermination controls, RNA processing (capping, polyadenylation, splicing), DNA methylation, heterochromatization, General and specific transcription factors, regulatory elements and mechanism of transcription regulation, transcriptional and post-transcriptional gene silencing, environmental regulation of gene expression.

# Unit-4:

Translation: Genetic code, Prokaryotic and Eukaryotic translation, mechanisms for initiation, elongation and termination, regulation of translation, co-and post- translational modifications of proteins. Homologous Recombination and Site-specific recombination.

- Molecular Biology of the Gene (4<sup>th</sup> Edition) *J.D. Watson, N.H. Hopkins, J.W. Roberts, J.A. Steitz and A.M Weiner*, The Benjamin/ Cummings Publ. Co. Inc, California
- Molecular Biology of the cell (3<sup>rd</sup> Edition) by *Bruce Alberts, Dennis Bray, Julian Lewis, martin Raff, Keith Roberts and James D. Watson,* Garland Publishing, Inc, New York & London
- Gene Cloning and DNA Analysis (4<sup>th</sup> Edition) by *T.A Brown*, Blackwell Science
- Essential of Molecular Biology (3<sup>rd</sup> edition) by *G.M. Malacinski& D. Freifelder*, Jones & Bartlett Publisher

BT-503: BIOCHEMISTRY	4+0
THEORY	
(Core)	

# Unit 1

**Amino acids and proteins:** Classification, structure and properties of amino acids; primary, secondary, tertiary, quaternary and domain structure of proteins, forces stabilizing protein structure, Ramachandran plot, DNA-protein and protein-protein interactions, protein folding, protein misfolding and related diseases; protein sequencing.

**Nucleic acids:** Primary and secondary structure of nucleic acids, Watson-Crick model of DNA, structural polymorphism of DNA and RNA, three dimensional structure of RNA, biosynthesis of purines and pyrimidines.

#### Unit 2

**Carbohydrates:** Classification and structure of carbohydrates, polysaccharides, glycoproteins and peptidoglycans, glycolysis, TCA cycle, oxidative phosphorylation, glycogen synthesis and breakdown, gluconeogenesis, interconversion of pentoses and hexoses. **Lipids** :Classification, structure and functions, biosynthesis of fatty acids, oxidation of lipids, triglycerides, phospholipids, sterols.

#### Unit 3

**Biophysical techniques:** Principles and applications of thin layer chromatography, gas chromatography, HPLC and FPLC, Principles and applications of X-ray diffraction, NMR, ESR, ORD/CD, fluorescence, UV, IR, visible and mass spectroscopy.

#### Unit 4

**Principles and applications of tracer techniques in biology:** Effect of radiation on biological systems, radioactive isotopes and their half life, autoradiography, radiation dosimetry, Cerenkov radiation, liquid scintillation spectrometry.

- Biochemistry by Stryer, Freeman publisher
- Biochemistry, Vol I, II, III by GeofferyZubey, WCB press
- Fundamentals of Biochemistry by Voet, Voet& Pratt, John Wiley publisher
- Principles of biochemistry by Albert Lehninger, David L Nelson & Michael M Cox, Mac Milan worth publisher.

BT-504: CELL BIOLOGY	4+0
THEORY	
(Core)	

Structure of prokaryotic and eukaryotic cells, Cellular organelles: Plasma membrane, cell wall, cytoskeleton- their structural organization; Mitochondria; Chloroplast; Nucleus and other organelles and their organization and function, genetic constitution of mitochondria and chloroplast, artificial membrane Liposomes.

# Unit 2:

Microscopic techniques: Principles and application of light, phase contrast, fluorescence, confocal, scanning and transmission electron microscopy, cytophotometry and flow cytometry, fixation and staining, Fluorescence *in-situ* hybridization (FISH), GISH (Genomic *in-situ* hybridization).

#### Unit 3:

Transport of nutrients, ions and macromolecules across membranes, Cell cycle: Mitosis, meiosis, role of cyclins and cyclin dependent kinases, regulation of Cdk-cyclin activity, Cdk inhibitors, induction of cancer with respect to cell cycle, molecular events and regulation in model systems, cell surface receptors, second messenger system, MAP kinase pathways, mechanism of signal transduction pathway.

#### Unit 4:

Molecular biology and biochemistry of cancer, oncogenes, tumor suppressor genes, chemical carcinogenesis, Cellular basis of differentiation and development- cell division, gametogenesis and fertilization, differential gene activity and cell differentiation, Morphogenetic determinants in egg cytoplasm, genetic regulation of early embryonic development in Drosophila, homeotic genes.

- Essential Cell Biology by *Bruce Albertset.al.*, Garland Publisher.
- Cell and Molecular Biology by F. D P deRobertes, LW & W Publisher.
- Molecular Biology of the Cell by *Alberts, Bray, Lewis, Raff, Roberts and Watson,* Garland Publishers.
- Molecular Cell Biology by H. Lodish, D. Baltimore, A. Bark, S. L. Zipursky, P. Matsudaira and J. Darnell, Scientific American Books.

# BT-505: LABORATORY COURSE-I 0+8 (Core) (PRACTICALS BASED ON COURSES BT 501-BT504)

- Cleanliness, media preparation, sterilization, culturing methods, dilution techniques.
- Staining techniques in microbiology; simple staining, gram staining, spore staining capsule staining, flagella staining.
- Isolation of pure culture by different techniques.
- Replica plating technique.
- Transformation of *E.coli* host strain with provided plasmid DNA using CaCl<sub>2</sub> method.
- Minipreparation of plasmid DNA by Alkaline Lysis method.
- Quantitative and qualitative estimation of DNA by Agarose gel electrophoresis.
- Isolation of genomic DNA from plant, fungal and bacterial sources.
- Determining the purity of isolated genomic/plasmid DNA using UV-Vis spectrophotometer.
- Isolation of protein from the given sample.
- Estimation of protein by Lowry-Folin method.
- Estimation of protein by Biuret method.
- Separation of protein by SDS-PAGE.
- Quantification of total sugar by Anthron method.
- Identification of reducing and nonreducing sugars.
- Blood group identification of M.Sc. students.

BT-506: SEMINAR (Core)	0+1
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# Semester 2

BT-507: RECOMBINANT DNA TECHNOLOGY	4+0
THEORY	
(Core)	

#### Unit 1:

Molecular tools and their applications: Restriction endonucleases, polymerases, nucleases, kinases, topoisomerases, gyrases, methylases and ligases. Cloning vectors: Plasmids, Bacteriophages, Cosmids, Phagemids, Artifical chromosomes (BAC, PAC, MAC).

#### Unit 2:

Construction and screening of genomic and cDNA libraries, EMSA (Electrophoretic mobility shift assay), DNA footprinting, Primer extension, SI mapping, RNase protection assay, Reporter assays, Principles and techniques of nucleic acid hybridization, Southern, Northern and Western hybridization/blotting, DNA microarray-fabrications, variations and applications, Serial Analysis of Gene Expression (SAGE).

#### Unit 3

Polymerase chain reaction: principle, different ingredients of PCR, primer-designing, variations-standard PCR, Touch down PCR, Hot- start PCR, Asymmetric PCR, Inverse PCR, Long PCR, High Fidelity PCR, Multiplex PCR, Nested PCR, Reverse transcriptase PCR, Real Time quantitative PCR, Applications of PCR in different fields.

#### Unit 4

Expression strategies for heterologous genes: vector engineering, codon optimization, host engineering, expression in bacteria, yeast, insects, mammalian cells and plants, *in-vitro* transcription and translation, T-DNA and transposon tagging.

- iGenetics by Peter J Russell, Benjamin/ Cummings, New York
- From Genes to Clones: Introduction to gene technology, by *Ernst-L Winnacker*, VCH Publication, Germany
- Principles of Gene Manipulation: An Introduction to genetic Engineering (6th Edition) by R.W. *Old and S.B. Primrose*, Blackwell Publication
- Genes IX by *Benjamin Lewin*, Oxford University Press, U.K.

<b>BT-508: ENZYME TECHNOLOGY</b>	4+0
THEORY	
(Core)	

#### Unit 1

Nomenclature and classification of enzymes, general properties of enzymes, active sites, cofactors and specificity.

Isolation, purification and large scale production of enzymes with principles and applications of the involved techniques, viz gel filtration, ion exchange and affinity chromatography, centrifugation and electrophoretic techniques.

#### Unit 2

Enzyme kinetics: Enzymatic reaction mechanisms, Michaelis-Menten equation, Effect of substrate, pH, temperature and inhibitors on enzyme activity.

Mechanism of enzyme action and regulation: Active and regulatory sites, chemical modification, feedback inhibition, positive and negative cooperativity, allosteric enzymes.

#### Unit 3

Isozymes, multienzyme complexes, artificial enzymes, catalytic antibodies. Enzyme engineering-strategies, directed evolution, degradation of unnatural substrates.

#### Unit 4

Industrial enzymes: In detergent, food, leather, dairy, medicines and chemical industries. Enzyme immobilization: Introduction, methods, applications and limitations.

- Enzymes: Biochemistry, Clinical Chemistry by T. Palmor, Harwood press
- Fundamentals of Enzymology: The cell and molecular biology of catalytic proteins, by NC Price and Steven, Oxford press.
- Biochemistry, Vol I, II, III by GeofferyZubey, WCB press
- Fundamentals of Biochemistry by Voet, Voet& Pratt, John Wiley publisher

BT-509: IMMUNOLOGY	4+0
THEORY	
(Core)	

Introduction: Phylogeny of Immune System, Innate and acquired immunity, Clonal nature of immune response, Primary and secondary immune response, Organization and structure of lymphoid organs, Cells of the immune system:Haematopoesis and differentiation, lymphocyte trafficking, B lymphocytes, T- lymphocytes, Macrophages, dendritic cells, natural killer and lymphokine activated killer cells, Eosinophils, Neutrophils and mast cells.Antigens and superantigens.Structure and function of immunoglobulins.

# Unit 2:

Major histocompatibility complex; Antigen processing and presentation, BCR and TCR, generation of immunological diversity, Complement system.Cell- mediated cytotoxicity : Mechanism of T cell and NK cell mediated lysis, antibody dependent cell mediated cytotoxicity, macrophage mediated cytotoxicity, effector mechanism.

#### Unit 3:

Regulation of immune response:Generation of humoral and cell medicated immune responses, Activation of B-and T-lymphoeytes, cytokines and their role in immune regulation, Immunological tolerance, Genetic control of immune responses. Immunoprophylatic intervention: Basic concepts of vaccination and different types of vaccines.

#### Unit 4:

Hypersensitivity, Autoimmunity, Tumor immunology, AIDS and other immunedeficiencies.Antigen and antibody interactions, Immunodiffusion, Immunoelectrophoresis, RIA, ELISA, Hybridoma technology and monoclonal antibodies.

- Cellular and Molecular Immunology by *Abbas et al.*, Saunder Publication.
- Essential Immunology by *Roitt*, Blackwell Publisher.
- Immunology by *Kuby*, Freeman Publisher.
- Immunology-a short course by *Benjamini*, Wiley-Liss Publisher.

BT-510: BIOINFORMATICS,	4+0
<b>BIOSTATISTICS AND GENOMICS THEORY</b>	
(Core)	

Introduction to Bioinformatics, use of Internet and search engines (WWW, HTML, URLs, Netscape, Explorer, Google, PUBMED), database management system, database browsing, data retrieval, sequence and genome database, databases such as GenBank, EMBL, DDBJ, Swissprot, PIR, TIGR, TAIR, Searching for sequence database like FASTA and BLAST algorithm, multiple sequence alignment, phylogenetic analysis and detection of open reading frames (ORFs).

#### Unit 2:

Molecular evolution and phylogenetic tree, Gene predictions, Introduction to computational structural biology, *in-silico* methods for structural predictions, Homology threading and modeling, *ab-initio* modelling; Validation of *in-silico* determined 3D structures of proteins, Computer aided drug design-tools and applications.

#### Unit 3:

Selection of sample or sampling, theory: qualitative, random and non-random sample. Collection of data, their classification, tabulation, graphic representation and diagrammatic representation, measures of central tendency and dispersion: mean, median, mode, range, standard deviations, variance, idea of two types of errors and level of significance, test of significance (F & I test); chi-square tests, sample linear regression and correlation.

#### Unit 4:

Introduction to science of omics-genomics, proteomics, metabolomics, transcriptomics, comparative genomics, nutrigenomics, lipidomics, cytomics, toxicogenomics, pharmacogenomics; whole genome sequencing strategies, first, second, third and fourth generation sequencing technologies, genome mapping-physical and genetic mapping techniques.

- Introduction to Bioinformatics by *Stephen A Krawetz and David D. Womble*, Humana Press.
- Bioinformatics: Sequence and Genome Analysis by *David W. Mount*, Cold Spring Harbor Laboratory Press
- Fundamental of Biostatistics (5<sup>th</sup> edition) by *Bernard Rosner*, Duxbury Thomson Learning.
- Basic Statistics (2<sup>nd</sup> edition) by *B. L. Agrawal*, Wiley Eastern India.
- Introductory Statistics for Biology Students by *T. A Hall,* Chapman & Hall publisher.

- Statistical Methods in Biology by *N. T. J Bailey*, Cambridge Press.
- Genomes by T.A. Brown, John Wiley & Sons Ltd, New York
- Genome analysis (Volume I, II, III and IV) a Laboratory Manual by *Bruce Birren, Eric D. Green, Sue Klapholz, Richard M. Myers and Jane Roskams,* Cold Spring Harbor Laboratory Press.

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# BT-511: LABORATORY COURSE-II 0+8 (Core) (PRACTICALS BASED ON COURSES BT-507-510)

- PCR amplification using different template DNA and different sets of primers.
- Analysis of PCR amplified DNA using Agarose gel Electrophoresis.
- Gel elution of expected size PCR amplicons.
- Cloning of gel eluted amplicons in suitable vector.
- Analysis of cloned product by Restriction analysis.
- Isolation of enzyme from the given sample.
- Purification of enzyme from the crude extract.
- Effect of substrate concentration on enzyme activity.
- Effect of temperature on enzyme activity.
- Effect of pH on enzyme activity.
- Effect of metals and chemical reagents on enzyme activity.
- Performing enzyme immobilization by Calcium alginate entrapment method.
- Search for different web-based biological databases.
- Retrieval of nucleotide /protein sequences from different databases.
- BLAST analysis of provided sequences.
- Multiple sequence alignment of retrieved sequences using suitable *in-silico* tools.
- Construction of phylogenetic tree by different methods (NJ method, Maximum Likelihood, etc.)
- Motif analysis using *in-silico* tools.
- Physio-chemical characterization of sequences using *in-silico* tools.
- Gene structure and chromosomal localization of genes using in-silico tools.
- Homology based 3D structural determination and validation of given protein.
- I-TASSER based 3D structural determination and validation of given protein.
- Genome wide *in-silico* prediction and identification of transcription factor gene families.
- Primer designing using different *in-silico* tools.
- Solving problems related with mean, median, mode, range, standard deviations, variance etc.
- Display of lymphoid organs in rat/mouse.
- To perform ouchterlony double immunodiffusion and analysis of antigen-antibody precipitation pattern.
- To perform radioimmunoassay to measure the concentration of antigen by using antibodies.
- To perform ELISA for quantitative and qualitative estimation of antigens.
- To perform western blotting for qualitative estimation of protein.

0+1

# Semester-3

BT-513: PLANT BIOTECHNOLOGY	4+0
THEORY	
(Core)	

#### Unit 1:

History of plant cell and tissue culture; Culture media; various types of culture; callus, suspension, nurse, root, meristem, etc.; *In vitro* differentiation; organogenesis and somatic embryogenesis. Micropropagation; Anther and microspore culture; Somaclonal variation; *In vitro* fertilization; *In vitro* germplasm conservation; Production of secondary metabolites; Synthetic seeds.

#### Unit 2:

Embryo culture and embryo rescue; Protoplast isolation, culture and fusion; selection of hybrid cells and regeneration of hybrid plants; symmetric and asymmetric hybrids, cybrids. Conventional versus non-conventional methods for crop improvement; Present status and recent developments on available molecular markers, transformation and genomic tools for crop improvements. Molecular marker-aided breeding, QTL, molecular marker assisted selection.

#### Unit 3:

Plant transformation technology: *Agrobacterium* mediated, Particle bombardment, Electroporation; transgene stability and gene silencing. Chloroplast Transformation, Genetic engineering for resistance against abiotic (drought, salinity, flooding, temperature, etc.) and biotic (insect pest, fungal, viral and bacterial diseases, weeds, etc.) stresses; Genetic engineering for increasing crop productivity by manipulation of photosynthesis, nitrogen fixation and nutrient uptake efficiency; Genetic engineering for quality improvement (protein, essential amino acids, vitamins, minerals nutrients, etc.) etc.

#### Unit 4:

Metabolic Engineering and Industrial Products: Plant secondary metabolites, control mechanisms and mainipulation of phenylpropanoid pathway, shikimate pathway; alkaloids, biodegradable plastics, therapeutic proteins, edible vaccines, purification strategies.

- Plant Tissue Culture: Application and Limitation by S. S. Bhojwani and M. K. Razdan, Elsevier Publication
- Plants, Genes and Agriculture by *Maarten J Chrispeels and David E. Sadava*, Jones & Bartlett Publishers

- An Introduction to Plant Tissue Culture by *M. K. Razdan*, Oxford & IBH Publishing Co. Pvt. Ltd.
- Plant Biotechnology: The genetic manipulation of plants by Adrian Slater, Nigel Scott, and Mark Fowler, Oxford University Press

<b>BT-514: ANIMAL BIOTECHNOLOGY</b>	4+0
THEORY	
(Core)	

#### Unit 1

Introduction to animal cell and tissue culture, its advantages and limitations, Applications of animal cell and tissue culture.

Basic techniques in animal cell culture: Disaggregation of tissue and setting up of primary culture, established cell line cultures, maintenance of cell culture, culture media and role of serum in cell culture, organ culture.

#### Unit 2

Biology and characterization of the cultured cells, measurement of growth, measurement of viability and cytotoxicity.

Scale up of animal cell culture, cell cloning, cell synchronization and transformation.

#### Unit 3

Stem cell cultures: Embryonic and adult stem cells, their isolation, culture and applications, animal cloning.

Transgenic animals: Construction of transgenic animals, gene knockouts, ethical and biosafety considerations.

#### Unit 4

Gene therapy: Genetic disorders, vector engineering, types of gene therapy, strategies of gene delivery, targeted gene replacement/augmentation, gene editing, gene correction, gene silencing.

Molecular markers linked to disease resistance genes, Application of RFLP in forensic, disease prognosis, genetic counseling and pedigree analysis.

- Animal Cell Culture: A practical approach by R.I. Freshney, IRL press.
- Culture of animal cells: A manual of basic techniques by R.I. Freshney, Willy-Liss and Sons publication.
- Animal cell culture technique by Martin Clynes, Springer publication.

<b>BT-515: BIOPROCESS TECHNOLOGY</b>	4+0
THEORY	
(Core)	

Introduction to bioprocess technology, bioreactors, Isolation, preservation and maintenance of industrial microorganisms, kinetics of microbial growth and death, media for industrial fermentation, air and media sterilization.

#### Unit 2:

Types of fermentation processes: Analysis of batch, fed-batch, and continuous bioreactors, stability of microbial reactors, analysis of mixed microbial populations, specialized bioreactors (pulsed, photobioreactors etc.), measurement and control of bioprocess parameters.

#### Unit 3:

Downstream processing: Introduction, removal of microbial cells and solid matter, foam separation, precipitation, filtration, centrifugation, cell disruptions, liquid-liquid extraction, chromatography, Membrance process, Drying and crystallization, Whole cell immobilization and its industrial application.

#### Unit 4:

Industrial production of chemicals: Alcohol (ethanol), acids (citric, acetic and gluconic), Solvents (glycerol, acetone, butanol), Antibiotics (penicillin, streptomycin, tetracycline). Amino acids (lysine, glutamic acid), Single cell protein, Use of microbes in mineral beneficiation and oil recovery, Introduction to food technology:Elementary idea of canning and packing, Sterilization and pasteurization of food products.,Technology of typical food/ food products (bread, cheese, idli), Food preservation.

- Principles of fermentation technology by *PF Stanbury, A Whitekar and SJ Hall*, Aditya Books.
- Bioprocess Engineering; Basic Concept by *ML Suler*& *F Kargi*, PHI Press.
- Operational modes of bioreactors (BIOTAL Series), Butterworh Heineman.
- A Textbook of Industrial Microbiology by *W Cruger&ACruger*, W. H Freeman (Panima) Publisher.

BT-516: ENVIRONMENTAL	4+0
BIOTECHNOLOGY	
THEORY	
(Core)	

Introduction to Environmental Science: Environmental Pollution: Classification of pollutants, Ecosystem structure and functions, abiotic and biotic component, Energy flow, food chain, food web, Ecological Pyramids-types, biogeochemical cycles. Air, Water, Soil, Noise and Thermal pollution: Their source, Effect and biotechnology based control measures. Solid waste pollution and its management.

#### Unit 2:

Waste water Treatment: Biological treatment system (Oxidative ponds, aerobic and anaerobic ponds, facultative ponds, aerated ponds), Biological waste treatment, activated sludge treatment, microbialpollution in activated sludge, percolating filters, waste water treatment by biofilms. Treatment scheme of Dairy, Distillery, Tannery, Sugar, Fertilizers, Refinery, Chemical and Antibiotic waste.

#### Unit 3:

Bioremediation & Phytoremediation: Biofeasibility, applications of bioremediation, Bioreduction, Phytoremediation. Microbial Leaching and biomining, Recovery of metals from solutions, Microbes in petroleum extraction, Microbial desulfurization of coal, microbial transportation of toxic metals, Biodegradation of chlorinated hydrocarbons and xenobiotic compounds, pesticides, oil spills, and toxic dyes industrial effluents.

#### Unit 4:

Biofertilizers, biopestisides and Integrated pest management (IPM). Energy & Biofuels: Nonconventional or renewable sources of energy, Energy from Biomass, Biosensors and biochips. Ozone depletion, UV-B, Green-house effect and acid rain, their impact andbiotechnological approaches for management.

- Biotechnology Expanding Horizons by *B.D. Singh*. 2<sup>nd</sup> Edition Kalyani Publishers.
- Microbial Ecology: Fundamentals & Applications by *Atlas, R.M.*Wc Brown.
- Environmental Microbiology by *A.H. Varman*, ASM Press.
- Biodegradation and Bioremediation by *Alexandar, M*. Wiley International.

# BT-517: INTRODUCTORY BIOTECHNOLOGY THEORY (Elective for other Department)

4+0

# Unit 1:

Biotechnology: An overview-definition, scope and importance of Biotechnology, Concept of Recombinant DNA technology and Gene Cloning. Microbial Biotechnology: A brief account of microbes in industry and agriculture, Metabolic engineering for over production of metabolites.

# Unit 2:

Plant Biotechnology: Introduction to plant tissue culture and its applications, Gene transfer methods in plants, Transgenic plants (A brief introduction), Chloroplast and mitochondria engineering. Introduction to animal cell and tissue culture and its applications, production of transgenic animals, cell transformation and cell lines, animal cloning.

# Unit 3:

Medical Biotechnology: (A brief account) Biotechnology in medicine, Vaccines, Diagnostic, Forensic, Gene therapy, Nano Medicine & Drug Delivery Cell & Tissue Engineering, Stem Cell therapy. Environmental Biotechnology: (A brief account) Role of biotechnology in pollution control, Sewage treatment, Energy management, Bioremediation, Restoration of degraded lands and Conservation of biodiversity.

# Unit 4:

Bioinformatics: (A brief account) Importance, Scope of Bioinformatics, world wide web as a tool, Bioinformatics institutes and databases, Bioinformatics training & limitations. Bio-business and Bio-safety, Biotechnology for developing countries and IPR

- Das H.K. (2004), Textbook of Biotechnology, Willey Dreamtech. Pvt. Ltd, New Delhi.
- Kumar H.D. (2004), A Text Book of Biotechnology, Eastern Willey Press, New Delhi.
- Gupta P.K. (2010), Biotechnology & Genomics, 5th Reprint, Rastogi Publications Meerut.
  8.
- Biotechnology Expanding Horizons by B.D. Singh. 2<sup>nd</sup> Edition Kalyani Publishers .
- Black J.G (2008) Microbiology- Principles and Explorations, 7th edition, John Wiley & Sons

# BT-518: LABORATORY COURSE-III (Core) (PRACTICALS BASED ON COURSES MBT513-516)

- Plant tissue culture lab set up.
- Media preparation and sterilization.
- Explant sterilization.
- Callus induction of provided explants in suitable media.
- Regeneration of explants using different media.
- Setting up of animal tissue culture laboratory.
- Introduction to the equipments used in animal tissue culture laboratory.
- Isolation of macrophages from mouse peritoneum and their culture.
- Primary culture of spleenocytes isolated from mouse.
- Cell counting using nauber's chamber/hemocytometer.
- Cell viability testing using trypan blue dye.
- Cell cloning by dilution method.
- Sub culturing/splitting of monolayer culture.
- Isolation of metagenomic DNA from soil samples/water samples.
- PCR amplification of metagenomic DNA with 16S/18S primers/ gene specific primers.
- Detection of coliforms for determination of the purity of potable water.
- Determination of total dissolved solids of water
- Determination of Hardness and alkalinity of water sample.
- Determination of dissolved oxygen concentration of water sample
- Determination of biological oxygen demand of sewage sample
- Determination of chemical oxygen demand (COD) of sewage sample.
- Production of enzymes by solid state fermentation (SSF) and submerged fermentation (SmF).
- Characterization of enzymes produced by SSF/SmF (pH optima, pH stability, temperature optima & stability, effect of metal ions, inhibitors)
- Ammonium sulfate precipitation.
- Purification of enzymes by chromatography techniques.

BT-519: SEMINAR (Core)	0+1
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# Semester-4

<b>BT-520: GENOMICS FOR CROP</b>	4+0
IMPROVEMENT	
THEORY	
(Elective)	

#### Unit 1:

Introduction to science of omics for crop improvement, Introduction to the plant genomenuclear, chloroplast and mitochondrial genomes, genome size and complexity, mapping of genome: genetic and physical maps, map-based cloning, molecular markers in plant genome analysis; RFLP, RAPD, STS, Microsatellite, SCAR (Sequence characterized amplified regions), SSCP (single strand conformational Polymorphism), and AFLP analysis, FISH and GISH for genome analysis.

#### Unit 2:

Plant gene expression and regulation, functional genomics-expression analysis using microarrays, transposon tagging and Insertional mutagenesis - methods and significance, TILLING and EcoTILLING, Diversity Array Technology, transcriptomics.

#### Unit 3:

Whole genome analysis: Genome size, strategies for sequencing genome, ordered genomic libraries (Cosmid, YAC, BAC libraries), Genome sequencing in plants–Principles and Techniques; Next generation sequencing technologies, Applications of sequence information in plant genome analyses; Comparative genomics, Detection of Single Nucleotide Polymorphism; Role of transcriptomics, proteomics and metabolomics in linking genome and phenome.

#### Unit 4:

Marker assisted selection (MAS), Genomic assisted breeding approaches, Genomics andgenoinformatics for crop improvement; Integrating functional genomics information on agronomically/economically important traits in plant breeding, tagging of agronomically important traits, RNA interference in crop improvement.

- Genomes by *T.A. Brown*, John Wiley & Sons Ltd, New York
- Genome analysis (Volume I, II, III and IV) a Laboratory Manual by *Bruce Birren, Eric D. Green, Sue Klapholz, Richard M. Myers and Jane Roskams,* Cold Spring Harbor Laboratory Press.

• Discovery Genomics, Proteomics and Bioinformatics, *Campbell AM & Heyer L*, 2004, Pearson Education.

BT-521: PROTEOMICS AND	4+0
NANOBIOTECHNOLOGY	
THEORY	
(Elective)	

#### Unit 1

Proteomics technology: Gel electrophoresis of protein- SDS-PAGE, Native gel electrophoresis, zymography, identification and analysis of proteins by 2D analysis, mass spectrometry, MALDI-TOF, NMR and X-ray crystallography.

#### Unit 2

Differential display proteomics, protein-protein interactions, yeast two hybrid system and phage display, GFP and RFP, western blot, metabolic engineering.

#### Unit 3

Chemical, physical and biological properties of biomaterials and bioresponse, biomineralization, biosynthesis and properties of natural materials (protein, DNA & polysaccharides).

#### Unit 4

Preparation and characterization of nanoparticles : nanoparticular carrier system, micro and nano fluidics, drug and gene delivery system, microfabrication, chip technologies, biosensors, nano-imaging.

- Discovery genomics, proteomics and bioinformatics, Campbell AM & Heyer L, 2004, Pearson education.
- Methods in proteome and protein analysis, Kamp RM, 2004, Springer.
- Handbook of nanostructured biomaterials and their applications in Nanobiotechnology, Nalwa HS, 2005, American Scientific Publication.
- Nanobiotechnology, Niemeyer CM & Mirkin CA, 2005, Wiley Interscience.

BT-522: BIOSAFETY, IPR AND	4+0
BIOETHICS	
THEORY	
(Elective)	

Biosafety and risk assessment issues, regulatory framework, National biosafety policies and law, The Cartagena protocol on biosafety, WTO and other international agreements related to biosafety; Cross border movement of germplasm; Risk management issues-containment.

#### Unit 2:

General principles for the laboratory and environmental biosafety; health aspects; toxicology, allergenicity, antibiotic resistance etc. Impact on environment; gene flow in natural and artificial ecologies; Sources of gene escape, tolerance of target organisms, creation of superweeds/superviruses etc.

#### Unit 3:

Ecological aspects of GMOs and impact on biodiversity; Monitoring strategies and methods for detecting transgenics; Radiation safety and non-isotopic procedures; Benefits of transgenics to human health, society and the environment.

#### Unit 4:

The WTO and other international agreements; Intellectual properties, copyrights, trademarks, trade secret, patents, geographical indications, etc.; Protection of plant variety and farmers right act; Indian patent act and amendments, patent filing; Convention on biological diversity; Implications of intellectual property rights on the commercialization of biotechnology products.

#### Suggested Readings:

Singh BD, 2007. Biotechnology: Expanding Horizons. Kalyani http://patentoffice.nic.in

<b>BT-523: ANIMAL CELL AND TISSUE</b>	4+0
CULTURE	
THEORY	
(Elective)	

Introduction of Animal Cell and Tissue Culture, History of development of Animal cell culture techniques, Significance and Applications of tissue culture techniques.

# Unit 2:

Requirements in Animal Cell Culture, Equipments used in Cell culture, Culture vessels, Aseptic techniques, Culture media, designing of culture media, Serum free media development.

# Unit 3:

Primary culture, secondary culture, cell line, cryopreservation, contaminations, organotypic culture, Insect Cell Culture: An Overview, In vitro transformation of animal cells, Types of cell culture. Cell culture in vaccine production and drug/therapeutics development.

# Unit 4:

Cell cycle analysis and Synchronization of cultures, cancer studies using cell culture, production of hybridoma and monoclonal antibody production. Animal cloning, Therapeutic cloning, Tissue engineering, Knock out animals.

- Freshney, R.I : Culture of Animal cells , Wiley Publications , New York.
- Edi. Jhon R.W. Masters : Animal cell culture- practical approach , Oxford University press, Oxford.
- Ed. R. Basega : Cell growth and division : A practical approach , IRL press,Oxford University press, Oxford.
- Ed. Martin Clynes : Animal cell culture techniques , Springer- Verlag, New York.
- F.Grasveld, George V. Kallias: Transgenic Animals, Academic press, Sandiego, USA.
- Asok Mukhopadhyay: Animal cell technology, IK International publishing House, New Delhi.

BT-524: SEMINAR (Core)	0+1
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BT-525: DISSERTATION (Core)	0+12

Each Student will have to submit an allotted Dissertation, which would be based on research works and will submit a report on which Viva-Voce will be conducted.

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